

## Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 16 (2006) 4497-4503

## Cyclopentane-based human NK1 antagonists. Part 1: Discovery and initial SAR

Paul E. Finke,<sup>a,\*</sup> Laura C. Meurer,<sup>a</sup> Dorothy A. Levorse,<sup>a</sup> Sander G. Mills,<sup>a</sup> Malcolm MacCoss,<sup>a</sup> Sharon Sadowski,<sup>b</sup> Margaret A. Cascieri,<sup>b</sup> Kwei-Lan Tsao,<sup>c</sup> Gary G. Chicchi,<sup>c</sup> Joseph M. Metzger<sup>d</sup> and D. Euan MacIntyre<sup>d</sup>

<sup>a</sup>Department of Medicinal Chemistry, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA

<sup>b</sup>Department of Molecular Pharmacology, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA

<sup>c</sup>Department of Immunology, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA

<sup>d</sup>Department of Pharmacology, Merck Research Laboratories, PO Box 2000, Rahway, NJ 07065, USA

Received 27 April 2006; revised 8 June 2006; accepted 9 June 2006 Available online 7 July 2006

**Abstract**—An initial investigation of the novel cyclopentane scaffold **6** afforded low nanomolar human NK1 antagonists having enhanced water solubility properties compared to morpholine **1**. A synthesis of this cyclopentane scaffold, having three contiguous chiral centers, and the unexpected determination that the 1,2-*trans*-2,3-*trans*-ring stereochemistry, as opposed to the *cis*-ether/phenyl configuration of the known structures **1**–**5**, is optimal for this class of antagonist are described. © 2006 Elsevier Ltd. All rights reserved.

The neurokinin-1 receptor (NK1) is a member of the seven-transmembrane G-protein coupled family of receptors and is associated with sensory neurons in the periphery and specific areas of the CNS. The natural ligand for NK1 is the tachykinin peptide substance P (SP) which has been implicated in the pathophysiology of a diverse range of conditions including asthma, inflammatory bowel disease, pain, psoriasis, migraine, movement disorders, cystitis, schizophrenia, emesis, and anxiety/depression. To morpholine 1 (Aprepitant,  $IC_{50} = 0.09 \text{ nM})^2$  is the only currently marketed hNK1 antagonist and was approved by the FDA for use as an anti-emetic for chemotherapy-induced nausea and vomiting (CINV).3 During the course of this work, 1 was also being extensively investigated as an anti-depressant for major depressive disorder.<sup>4</sup> While 1 has been demonstrated in the clinic to have good bioavailability upon oral administration, a parenteral formulation was not possible due to its low solubility in appropriate aqueous vehicles (0.2 µg/mL in isotonic

Keywords: Neurokinin-1 receptor; NK1 antagonist; Cyclopentane-based structure.

saline, pH 8.2).<sup>5</sup> This limitation was initially addressed with the development of the aqueous soluble N-phosphoryl prodrug 2.<sup>5</sup> While increased solubility was also addressed with the development of the more soluble, fully basic derivative 4,<sup>6</sup> the identification of additional suitably soluble parent entities for both oral and parenteral use still remained an important need.<sup>7–9</sup>

As part of our ongoing pursuit of structurally diverse NK1 antagonists, the use of the 1,2-cis-2,3-trans-cyclohexane scaffold **5** (hNK1 IC<sub>50</sub> = 1.5 nM)<sup>10</sup> was previously investigated as a replacement for the core morpholine of structure **3** (hNK1 IC<sub>50</sub> = 0.09 and 1.1 nM for **3a** and **3b**)<sup>11</sup> from which **1** was later derived.<sup>2</sup> Herein, we report the initial synthesis of several isomers of the ring-contracted cyclopentane core structure **6**.<sup>7</sup> The structure–activity relationships (SAR) for this scaffold are discussed in terms of the stereochemistry at the three contiguous chiral centers and the enhanced water solubility achieved with this scaffold via incorporation of various basic moieties at C-3. Initial efforts first targeted derivatives based on our original morpholine structures **3a** and **3b**, having a 3,5-bis-trifluoromethylbenzyl ether at C-1 and an unsubstituted phenyl at C-2.<sup>12</sup> Subsequent efforts are detailed in the accompanying manuscript.<sup>13</sup>

<sup>\*</sup>Corresponding author. Tel.: +1 732 594 7456; fax: +1 732 594 5790; e-mail: paul\_finke@merck.com

The synthesis<sup>14</sup> of the core cyclopentane scaffold having an oxygen functionality at C-1, the required phenyl at C-2, and a functionalizable ester moiety at C-3 was based on the reported synthesis of 7 in 28% yield (Scheme 1).15 Subsequent to this work, an improved racemic route<sup>16</sup> to 7 and an asymmetric synthesis 17 of 9 were developed. The stereochemistry at the C-2 phenyl/C-3 ester was always the thermodynamically more stable trans-configuration.<sup>17,18</sup> Reduction of 7 with L-Selectride® produced exclusively the 1,2-cis-diastereomer 8, while use of sodium borohydride in methanol resulted in a chromatographically separable 1:3 mixture of predominantly the lower  $R_{\rm f}$  trans-isomer 9. This stereochemical selectivity allowed for a thorough evaluation of both alcohol isomers. The *trans*-assignment for **9** was confirmed by an <sup>1</sup>H NMR NOE experiment. <sup>19</sup> Separate hydroxyl alkylation of 8 and 9 with 3,5-bis-trifluoromethylbenzyl bromide and sodium hydride afforded the intermediate racemic ethers 10 and 11. Interestingly, the hNK1 affinity of derivatives prepared from the 1,2-trans-ester 11 was generally 3- to 5-fold greater than that of those derived from the 1,2-cis-ester 10 (see below); thus, the trans-series was preferentially investigated.

Hydrolysis of ester 11 gave acid 12 (Scheme 2) which was converted into amides 13. (Note that the *cis*-derivatives can be similarly prepared from ester 10.) Borane reduction then afforded the methylene-spaced amines 14 which could be alkylated to give a variety of derivatives (see Scheme 3). Alternatively, lithium borohydride reduction of 11 afforded the alcohol 15. Conversion to the bromide and alkylation of piperidine or morpholine afforded 16 and 17 as an alternate synthesis of aminomethylene derivatives. The ester 11 could also be converted to several *N*-cyclopentylamino derivatives via Curtius rearrangement of acid 12 and reaction of the intermediate isocyanate with either alcohols, to afford carbamates 18a–20a, or with amines, to afford ureas 21a–23a. Methylation and/or hydrogenation of the

CBz group of **19a** afforded the racemic *N*-cyclopentyl amines **24a** and **24b** which on alkylation (see Scheme 3) afforded analogs of structures **3** and **5**. Acylation of **24b** also afforded the N–Me derivatives **18b** and **23b**.

NC O-Et Et-O NC O-Et Me-O O-Me

O+ A NC 
$$(33\%)$$
 Me-O  $(33\%)$   $(57\%)$ 

**Scheme 1.** Reagents and conditions: (a) (i) piperidine, EtOH, 35–65 °C; (ii) NaCN, 35–80 °C, 1 h; (iii) ClCH<sub>2</sub>CH<sub>2</sub>CO<sub>2</sub>Et, 35–80 °C, 5 h; (b) 6 M HCl, reflux, 48 h; (c) MeOH, HCl (g), HC(OMe)<sub>3</sub>, 65 °C, 16 h; (d) NaOMe, MeOH or LiHMDS, THF; (e) L-Selectride<sup>®</sup>, THF (selective for **8**); (f) NaBH<sub>4</sub>, MeOH (1:3 ratio of **8:9**); (g) NaH, DMF.

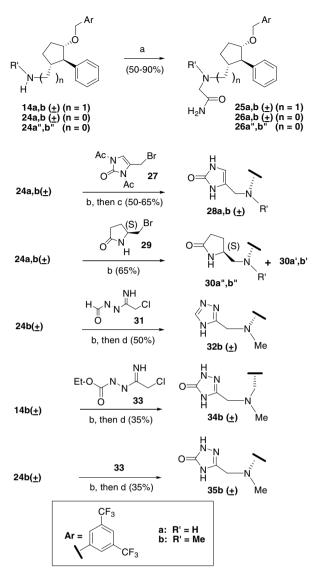
Scheme 2. Reagents and conditions: (a) NaOH, MeOH; (b) oxalyl chloride, DMF (cat), DCM; (c) NH<sub>3</sub> or MeNH<sub>2</sub> (aq, excess), THF; (d) BH<sub>3</sub>–Me<sub>2</sub>S, THF; (e) LiBH<sub>4</sub>, THF; (f) Ph<sub>3</sub>P–Br<sub>2</sub>, MeCN; (g) piperidine or morpholine (excess), MeCN; (h) NaN<sub>3</sub>, acetone/water, -10 °C; (i) PhMe, 85 °C; (j) MeOH, DIPEA, DMAP (cat); (k) BnOH or (S)-PhCH(Me)OH, DIPEA, DMAP (cat), PhMe, 85 °C; (l) NH<sub>3</sub>, MeNH<sub>2</sub> or Me<sub>2</sub>NH (aq, excess), dioxane; (m) H<sub>2</sub> (50 psi), 10% Pd/C, EtOH; (n) MeI, NaH, DMF; (o) CICO<sub>2</sub>Me, DIPEA, DCM; (p) CICONMe<sub>2</sub>, DIPEA, DCM. The 1,2-cis derivatives were similarly prepared from 10 and are designated as c (R' = H) and d (R' = Me) in the text and Table 1.

Alternatively, reaction of the above isocyanate with (S)α-methylbenzyl alcohol afforded 20a as a silica gel separable mixture of the diastereomeric CBz intermediates 20a' and 20a". The 1.2-cis-series also gave the two corresponding separable diastereomers; thus, all four enantiomerically enriched forms of 24 were available. The binding data (see below) were initially used to differentiate the more active and less active diastereomers. Based on modeling and by analogy to the known morpholine cis-stereochemistry, the stereochemistry for the active trans-enantiomer was tentatively assigned [1S,2S,3R] as shown for structure 24 in Scheme 2 and structure C in Table 1 (for  $R = NH_2$ ). The absolute assignment for the trans-series was later confirmed by single-crystal X-ray analysis of a resolved 4-fluorophenyl acid salt derivative of 9.12,13 The absolute stereochemistry of the active enantiomer in the cis-series was assumed to be the same as that of structure 3, i.e., [1S,2R,3S] as in structure **D**, Table 1 (for R = NH<sub>2</sub>).

Alkylation of the N–Me and N–H amines 14 and 24 (Scheme 3) afforded a variety of amine derivatives of varying basicity. Iodoacetamide afforded the glycinamide derivatives 25-26 as analogs of 3b, N,N-diacetyl bromomethylimidazolinone (27) gave 28 after removal of the N-acetyl groups, and use of (S)-bromomethylpyrrolidinone (29) allowed for the direct separation of diastereomeric lactams 30 (lower  $R_f$  diastereomers 30a" and 30b" being derived from the more potent enantiomer). Alternatively, the N–Me derivatives could also be alkylated with N-formyl- (31) or N-(methoxycarbonyl)-2-chloroacetamidrazone (33) followed by cyclization

at 150 °C in xylenes to afford the triazole **32b** and triazolinones **34b** and **35b**, which were analogous to the triazolinone morpholine structure **3a**. <sup>2,11</sup>

While non-basic morpholine 1 has been found to have good bioavailability upon oral administration to several species (e.g., rat, F = 53%; dog, F = 40%), formulation for parenteral use was not possible due to its low solubility in appropriate aqueous vehicles (0.2 µg/mL in isotonic saline, final pH 8.2).5 The possibility of enhanced water solubility was expected for this more basic cyclopentane scaffold, since an exocyclic nitrogen would no longer be benzylic nor part of a morpholine ring. Both of these features greatly diminish the basicity of the morpholine nitrogen (p $K_a$  of 3a < 3.5 in 1:1 methanol/ water). 11,20 Indeed, as listed in Table 1, the water solubility at pH 5 (0.1 M sodium acetate/acetic acid buffer) for many of these derivatives was found to be significantly enhanced over that of 1 (<0.0005 mg/mL at pH 5). 21 For the N-H glycinamides 25a and 26a (0.59 and 1.1 mg/mL) the solubility enhancement was over 1000fold, while the solubility for the corresponding N-Me analogs **25b** and **26b** (0.30 and 0.016 mg/mL) was less impressive. Good solubilities were also seen for 1,2trans-heterocyclic derivatives 28b, 30a", and 30b" (0.45, 4.0, and 0.51 mg/mL, respectively) with the N-Me triazolinone 35b (0.20 mg/mL) giving at least a 400-fold increase. In this case, the corresponding less active 1,2cis-triazolinone 35d (see Table 1) was also slightly less soluble (0.08 mg/mL). These enhanced solubilities are most likely due to the increased basicity of these derivatives. For example, the trans-derivatives 26a and 26b



Scheme 3. Reagents and conditions: (a)  $ICH_2CONH_2$ , DIPEA,  $CH_3CN$ ; (b) DIPEA,  $CH_3CN$  (for 29, 90 °C in sealed vial); (c) MeNH<sub>2</sub>, THF; (d) xylenes 150 °C. The 1,2-*cis* derivatives in Table 1 were similarly prepared from *cis*-24 and are designated  $\mathbf{c}$  ( $\mathbf{R}' = \mathbf{H}$ ) and  $\mathbf{d}$  ( $\mathbf{R}' = \mathbf{M}\mathbf{e}$ ).

had measured p $K_a$ s of 6.6 and 5.8 in 1:1 methanol/water.<sup>20</sup>

The hNK1 binding affinities for both the neutral and basic target compounds, as well as several of the intermediates, were determined by measuring their ability to displace [125I]SP from human NK1 receptor stably expressed in CHO cells.<sup>22</sup> The hNK1 binding results are summarized in Table 1 with significant binding affinity being observed in several classes of compounds. The initial cis-derivatives 18c and 21c indicated some hNK1 affinity (hNK1 IC<sub>50</sub> = 5.3 and 9.8 nM), however, unexpectedly the corresponding trans-isomers 18a and 21a actually showed enhanced affinity (IC<sub>50</sub> = 0.89 and 3.5 nM). Also, other neutral trans-nitrogen derivatives (i.e., 13a, 18b, 22a, and 23a;  $IC_{50} = 7.2$ , 6.2, 2.7, and 5.0 nM, respectively) and even the alcohol intermediate 15  $(IC_{50} = 0.77 \text{ nM})$  achieved moderate potency, although the acid 12 had poor hNK1 affinity. While pre-

vious studies in the six-membered core piperidine9b and morpholine<sup>11</sup> scaffolds had reported that the cis-configuration afforded significantly better potency than the trans-configuration ( $\sim$ 200-fold for the [2S,3S] morpholine 3a and its trans [2S,3R] isomer), in this cyclopentane scaffold the opposite relationship was found. With the possible exceptions of 24a and 24c and 26b and 26d the trans-derivatives generally afforded a 2- to 5-fold enhancement over the cis-configuration. More importantly, although the racemic 1,2-cis-N-H glycinamide 26c indicated poorer binding affinity compared to 5<sup>10</sup> and the equivalent acetamide derivative  $3b^{11}$  (IC<sub>50</sub> = 8.3 nM vs 1.5 and 1.1 nM, respectively), the non-racemic [1S,2S,3R] trans N-H derivative 26a" was very comparable (IC<sub>50</sub> = 0.95 nM). At the time, potential use of the 1,2-trans-cyclohexane had not been considered based on the earlier morpholine results. Although the effect of an additional N-methyl to afford the tertiary amines was beneficial in the cis-series  $(IC_{50} = 8.3 \text{ vs } 2.1 \text{ nM for } 26c \text{ and } 26d)$ , the results in the more potent *trans*-series were mixed (IC<sub>50</sub> = 1.3 vs 2.7 nM for 26a and 26b; 5.2 vs 2.4 nM for 28a and **28b**; 1.5 vs 0.81 nM for **30a**" and **30b**"). Insertion of an additional methylene as in analogs 25a and 25b also made little difference in binding. Reversing the amide as in the (S)-lactams 30a" and 30b" gave comparable results to the glycinamides, although the corresponding (R)-lactams generally showed about 2-fold poorer affinity (data not shown). Unfortunately, the targeted imidazolinone, triazole, and triazolinone derivatives (28b, 32b, and 35b;  $IC_{50} = 2.4$ , 1.1, and 2.0 nM, respectively) did not show the anticipated 10-fold improvement obtained with the morpholines (IC<sub>50</sub> = 0.09 vs 1.1 nM for **3a** and **3b**). 11 As would be expected, there was an enantiomeric preference as seen with the chiral amines 24a' and 24a'' (IC<sub>50</sub> = 100 and 2.3 nM), glycinamides 26a' and 26a'' (IC<sub>50</sub> = 45 and 0.95 nM), and (S)-lactams 30a' and 30a'' ( $IC_{50} = 260$  and 1.5 nM).

This general enhanced affinity of the 1,2-trans-series over the 1,2-cis-series is likely due to the increased flexibility of the five- versus six-membered ring in which the 1,2-trans orientation of the two phenyl moieties can afford a more optimal hNK1 pharmacophore fit. Interestingly, the extent of this enhancement is also quite dependent on the C-3 substituent, ranging from a cis:trans IC<sub>50</sub> ratio of 0.8 for **26b** and **26d** to 6 for **26a** and **26c**. The nature of the C versus N linkage at C-3 might also be important, although the N-substituent would be expected to be similarly 'trans' to the phenyl. Thus, it is noteworthy that it is the relative 2,3-substituent configurations which are reversed while the ether absolute configuration is conserved. In addition, these results again demonstrate that potent hNK1 binding is not dependent on the presence of a fully basic nitrogen at the 3-position relative to the ether, in fact, the trans-amine 14a showed less hNK1 affinity than the alcohol 15. This toleration of a wide variety of functionality at the C-3 position suggests that the receptor binding domain in the region of the C-3 moiety is relatively unstructured, although there is a general preference for a polar, small to moderately sized moiety (see 16, 17, 19a, 23a and 23b having larger R' groups). However, the choice of substituent at the C-3 position was

Table 1. Structures, solubilities,  $pK_as$ , and hNK1 binding affinities of selected compounds

Compound	Stereochemistry	-R	NK1 $IC_{50}^{a}$ (nM)	$(SEM, n)^b$	Solubility at pH 5 (mg/mL) <sup>c</sup>	$pK_a^{d}$
1	(2R,3S)	_	0.09 <sup>e</sup>	_	< 0.0005	<3.5
3a	(2S,3S)	_	$0.09^{f}$		0.003	<3.5
3b	(2S,3S)	_	1.1 <sup>f</sup>	_		
5	(1S, 2S, 3R)	_	1.5 <sup>g</sup>			
12	trans, A/C	−CO <sub>2</sub> H	67	(n = 1)		
13a	trans, A/C	-CONH <sub>2</sub>	7.2	(1.2, 2)		
14a	trans, A/C	-CH <sub>2</sub> NH <sub>2</sub>	2.7	(0.3, 3)		
14b	trans, A/C	-CH <sub>2</sub> NHMe	1.9	(0.5,3)		
15	trans, A/C	−CH <sub>2</sub> OH	0.77	(0.16, 3)		
16	trans, A/C	-CH <sub>2</sub> -1-piperidinyl	5.0	(n = 1)		
17	trans, A/C	-CH <sub>2</sub> -4-morpholinyl	8.2	(0.9, 3)		
18a	trans, A/C	−NHCO <sub>2</sub> Me	0.89	(0.10, 4)		
18b	trans, A/C	$-NMeCO_2Me$	6.2	(1.0, 4)		
18c	cis, B/D	-NHCO <sub>2</sub> Me	5.3	(1.7, 3)		
19a	trans, A/C	−NHCO <sub>2</sub> Bn	40	(n = 1)		
19c	cis, B/D	−NHCO <sub>2</sub> Bn	300	(n = 1)		
21a	trans, A/C	-NHCONH <sub>2</sub>	3.5	(0.3, 3)		
21c	cis, B/D	-NHCONH <sub>2</sub>	9.8	(2.2, 3)		
22a	trans, A/C	-NHCONHMe	2.7	(0.3, 3)		
23a	trans, A/C	-NHCONMe <sub>2</sub>	5.0	(0.7, 3)		
23b	trans, A/C	-N(Me)CONMe <sub>2</sub>	16	(n = 1)		
24a	trans, A/C	$-NH_2$	5.7	(n=1)		
24b	trans, A/C	-NHMe	0.83	(0.38, 3)		
24c	cis, B/D	$-NH_2$	7.9	(2.4, 3)		
24d	cis, B/D	–NHMe	7.2	(n = 1)		
24a'	trans, A	$-NH_2$	100	(n=1)		
24a"	trans, C	$-NH_2$	2.3	(0.2, 3)		
25a	trans, A/C	-CH <sub>2</sub> NHCH <sub>2</sub> CONH <sub>2</sub>	1.9	(0.7, 3)	0.59	
25b	trans, A/C	-CH <sub>2</sub> NMeCH <sub>2</sub> CONH <sub>2</sub>	1.9	(0.3, 8)	0.30	
26a	trans, A/C	-NHCH <sub>2</sub> CONH <sub>2</sub>	1.3	(0.1, 7)	1.1	6.6
26b	trans, A/C	-NMeCH <sub>2</sub> CONH <sub>2</sub>	2.7	(0.4, 6)	0.016	5.8
26c	cis, B/D	-NHCH <sub>2</sub> CONH <sub>2</sub>	8.3	(2.0, 3)	0.80	
26d	cis, B/D	-NMeCH <sub>2</sub> CONH <sub>2</sub>	2.1	(0.3, 6)		
26a'	trans, A	-NHCH <sub>2</sub> CONH <sub>2</sub>	45	(n = 1)		
26a"	trans, C	-NHCH <sub>2</sub> CONH <sub>2</sub>	0.95	(0.06, 6)		
	,	Н				
<b>28a</b> $(R' = H)$	trans, A/C	N-	5.2	(0.4, 6)		
<b>28b</b> ( $R' = Me$ )	trans, A/C	O <del> </del>	2.4	(0.7, 3)	0.45	
200 (11 1110)	, 11 C	H R'		(0.7, 5)	01.12	
30a' (R' = H)	trans, A	O= (S) 2	260	(35, 3)		
30a'' (R' = H)	trans, C	N N	1.5	(0.3, 3)	4.0	
30b'' (R' = Me)	trans, C	H R'	0.81	(0.35, 3)	0.51	
32b	trans, A/C	N-N yw	1.1	(0.5, 3)		
~	,	H Me		(0.0, 0)		
<b>34b</b> ( <i>n</i> = 1)	trans, A/C	Me (/)n	4.9	(1.3, 3)	0.01	
<b>35b</b> $(n = 0)$	trans, A/C	H.N.N.N.	2.0	(0.5, 3)	0.20	
<b>35d</b> $(n = 0)$	cis, B/D		4.7	(n = 1)	0.08	
( 0)	J, 2.2	ŭ 'i		( 1)		

<sup>&</sup>lt;sup>a</sup> Displacement of <sup>125</sup>I-labeled SP from the cloned hNK1 receptor expressed in CHO cells. <sup>b</sup> Except as noted (n = 1), data are an average of 2–8 independent replicate titrations.<sup>22</sup> <sup>c</sup> Aqueous solubilities were measured in 0.1 M sodium acetate/acetic acid buffer at pH 5 (n = 2).<sup>21</sup> <sup>d</sup> Titrations were preformed in 1:1 methanol/water.<sup>20</sup>

e See Ref. 2.
f See Ref. 11.

g See Ref. 10.

found to be critical for in vivo potency and for obtaining the desired physical properties (also see Ref. 13).

In order to determine whether these new hNK1 antagonists could inhibit the action of SP in vivo, a previously described assay (SYVAL<sup>23</sup>) was utilized.<sup>2,11</sup> Intravenous administration of capsaicin or resiniferatoxin causes a dose-dependent vascular leakage in the esophagus, trachea, and bladder of guinea pigs and can be quantified with Evans Blue dye. This response is mediated by the endogenous release of SP from capsaicin-sensitive nerve fibers and can be inhibited by the systemic administration of NK1 receptor antagonists. Test compounds were administered orally at differing doses and time intervals before capsaicin or resiniferatoxin challenge, thus serving both as a functional NK1 inhibition readout as well as a pharmacokinetic measurement. The utility of these derivatives in this assay and the in vivo importance of the C-3 moiety were initially demonstrated with two of the above compounds. When the trans-compound 26a  $(IC_{50} = 1.3 \text{ nM})$  was administered orally at 1 mg/kg and 1 h prior to challenge, a 60% inhibition was achieved. However, when the neutral carbamate 18a  $(IC_{50} = 0.89 \text{ nM})$  was administered, <25% inhibition was observed.

The initial synthesis of a variety of 3-amino and 3-aminomethylene derivatives was investigated in this 1-benzyloxy-2-phenylcyclopentane scaffold based on the previous morpholine structure 3. The synthesis involved the stereoselective synthesis of either the 1,2-cis or 1,2trans-hydroxy intermediates 8 and 9. The 3-amino derivatives were also available in chiral form through separation of the diastereomeric (S)- $\alpha$ -methylbenzyl carbamates 20a and 20c. Several basic cyclopentane derivatives demonstrated the desired enhanced water solubility at pH 5 necessary for an intravenous formulation. The hNK1 binding affinity for this series of compounds was found to be relatively insensitive to the functionality at C-3, although moderately sized hydrophilic moieties were preferred. Preliminary results for the glycinamide derivative 26a indicated that this class of hNK1 antagonist was also capable of in vivo inhibition of SP-elicited systemic plasma extravasation after oral administration. The finding that the 1,2-trans-2,3-transconfiguration in this scaffold was significantly better than the preferred cis-arrangement in our previous morpholine core structures was unexpected. These results led to additional investigations of this scaffold as discussed further in the accompanying manuscript<sup>13</sup> elsewhere.24,25

## Acknowledgments

We thank George A. Doss for performing the <sup>1</sup>H NMR NOE experiment on compound **9** and Karen A. Owens for some of the aqueous solubility measurements.

## References and notes

(a) Quatara, L.; Maggi, C. A. Neuropeptides 1998, 32, 1;
 (b) Rupniak, N. M. J.; Kramer, M. S. Trends Pharmacol.

- Sci. 1999, 20, 485; (c) Dando, T. M.; Perry, C. M. Drugs 2004, 64, 777.
- Hale, J. J.; Mills, S. G.; MacCoss, M.; Finke, P. E.; Cascieri, M. A.; Sadowski, S.; Ber, E.; Chicchi, G. G.; Kurtz, M.; Metzger, J.; Eiermann, G.; Tsou, N. N.; Tattersall, F. D.; Rupniak, N. M. J.; Williams, A. R.; Rycroft, W.; Hargreaves, R.; MacIntyre, D. E. J. Med. Chem. 1998, 41, 4607.
- 3. Pendergrass, K.; Hargreaves, R.; Petty, K. J.; Carides, A. D.; Evans, J. K.; Horgan, K. J. *Drugs Today* **2004**, *40*, 853.
- Kramer, M. S.; Cutler, N.; Feighner, J.; Shivastava, R.; Carman, J.; Sramek, J. J.; Reines, S. A.; Liu, G.; Snavely, D.; Wyatt-Knowles, E.; Hale, J. J.; Mills, S. G.; MaCoss, M.; Swain, C. J.; Harrison, T.; Hill, R. G.; Hefti, F.; Scolnick, E. M.; Cascieri, M. A.; Chicchi, G. G.; Sadowski, S.; Williams, A. R.; Hewson, L.; Smith, D.; Carlson, E. J.; Hargreaves, R. J.; Rupniak, N. M. J. Science 1998, 281, 1640.
- Hale, J. J.; Mills, S. G.; MacCoss, M.; Dorn, C. P.; Finke, P. E.; Budhu, R. J.; Reamer, R. A.; Huskey, S. W.; Luffer-Atlas, D.; Dean, B. J.; McGowan, E. M.; Feeney, W. P.; Chiu, S. L.; Cascieri, M. A.; Chicchi, G. G.; Kurtz, M. M.; Sadowski, S.; Ber, E.; Tattersall, F. D.; Rupniak, N. M. J.; Williams, A. R.; Rycroft, W.; Hargreaves, R.; Metzger, J. M.; MacIntyre, D. E. J. Med. Chem. 2000, 43, 1234.
- Harrison, T.; Owens, A. P.; Williams, B. J.; Swain, C. J.; Williams, A.; Carlson, E. J.; Rycroft, W.; Tattersall, F. D.; Cascieri, M. A.; Chicchi, G. G.; Sadowski, S.; Rupniak, N. M. J.; Hargreaves, R. J. J. Med. Chem. 2001, 44, 4296.
- For a preliminary report on this work, see: Finke, P. E.; Meurer, L. C.; MacCoss, M.; Mills, S. G.; Sadowski, S.; Cascieri, M. A.; Metzger, J.; Eiermann, G.; MacIntyre, D. E. American Chemical Society 219th National Meeting, San Francisco, California, March 26–31, 2000.
- 8. Williams, B. J.; Teall, M.; McKenna, J.; Harrison, T.; Swain, C. J.; Cascieri, M. A.; Sadowski, S.; Strader, C.; Baker, R. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 1903.
- 9. (a) Harrison, T.; Williams, B. J.; Swain, C. J.; Ball, R. G. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 2545; (b) Harrison, T.; Owens, A. P.; Williams, B. J.; Swain, C. J.; Baker, R.; Hutson, P. H.; Sadowski, S.; Cascieri, M. A. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 209.
- Mills, S. G.; MacCoss, M.; Underwood, D.; Shah, S. K.; Finke, P. E.; Miller, D. J.; Budhu, R. J.; Cascieri, M. A.; Sadowski, S.; Strader, C. D. *Bioorg. Med. Chem. Lett.* 1995, 5, 1345.
- Hale, J. J.; Mills, S. G.; MacCoss, M.; Shah, S. K.; Qi, H.; Mathre, D. J.; Cascieri, M. A.; Sadowski, S.; Strader, C. D.; MacIntyre, D. E.; Metzger, J. M. J. Med. Chem. 1996, 39, 1760.
- Finke, P. E.; MacCoss, M.; Meurer, L. C.; Mills, S. G.; Caldwell, C. G.; Chen, P.; Durette, P. L.; Hale, J. J.; Holson, E.; Kopka, I.; Robichaud, A. US Patent 5,750,549, 1998; Chem. Abstr. 1997 127, 17433.
- Meurer, L. C.; Finke, P. E.; Owens, K. A.; Tsou, N. N.; Ball, R. G.; Mills, S. G.; MacCoss, M.; Sadowski, S.; Cascieri, M. A.; Chicchi, G. G.; Egger, L. A.; Luell, S.; Metzger, J. M.; MacIntyre, D. E.; Rupniak, N. M. J.; Williams, A. R.; Hargreaves, R. J. Bioorg. Med. Chem. Lett., 2006, 16 (following manuscript page).
- 14. Representative experimental details and spectral data have been described in Ref. 12. Indicated yields were not optimized. All mass spectral and <sup>1</sup>H NMR data were consistent with the assigned structures.
- 15. Baker, W.; Leeds, W. G. J. Chem. Soc. 1948, 974.
- 16. Desai, R. C.; Cicala, P.; Meurer, L. M.; Finke, P. E. *Tetrahedron Lett.* **2002**, *43*, 4569.

- Kuethe, J. T.; Wong, A.; Wu, J.; Davies, I. W.; Dormer, P. G.; Welch, C. J.; Hillier, M. C.; Hughes, D. L.; Reider, P. J. J. Org. Chem. 2002, 67, 5993.
- 18. In subsequent studies, minor amounts of 1,2-*trans*-2,3-*cis* compounds were isolated, but the hNK1 affinity was significantly diminished and, thus, these isomers were never pursued.
- 19. A <sup>1</sup>H NMR NOE experiment of the more polar isomer showed an unambiguous interaction between the C-1 and C-3 protons, thus, confirming the 1,2-trans-2,3-trans assignment for 9.

- 20. The reported  $pK_a$  values were obtained by titration in 1:1 methanol/water. The presence of 50% MeOH in a titration has the effect of lowering the  $pK_a$  of an amine between 0.3 and 0.9 U (with 0.5 U as a typical value) as compared to its value in water (i.e., the actual  $pK_a$  of **26b** in water is likely closer to 6.3). <sup>26</sup>
- 21. Solubilities were determined by HPLC after sonication in buffer at 1 mg/mL and filtration. The solubility for 1 and 3a was determined in 0.1 M potassium hydrogen phosphate buffer at pH 5, while the solubilities for selected

- cyclopentane compounds were done in  $0.1\,\mathrm{M}$  sodium acetate/acetic acid buffer adjusted to pH 5. The reported results are an average of n=2.
- Cascieri, M. A.; Ber, E.; Fong, T. M.; Sadowski, S.; Bansal, A.; Swain, C. J.; Seward, E. M.; Frances, B.; Burns, D.; Strader, C. D. Mol. Pharmacol. 1992, 42, 458.
- Acronym for resiniferatoxin-induced SYstemic VAscular Leakage.
- (a) Finke, P. E. National Medicinal Chemistry Symposium, Madison, WI. June 27–July 1, 2004; (b) Finke, P. E.; Meurer, L. C.; Mills, S. G.; MacCoss, M.; Qi, H. U.S. Patent 6,479,518, 2002. Chem. Abstr. 2002, 136, 279139x.
- 25. The use of the *trans* ether/phenyl configuration was subsequently applied to other six-<sup>27</sup> and seven-membered ring scaffolds.<sup>28</sup>
- Albert, A.; Serjeant, E. P. The Determination of Ionization Constants, 3rd ed.; Chapman and Hall: New York, NY, 1984, p 35.
- (a) Owens, S. N.; Seward, E. M.; Swain, C. J.; Williams, B. J. U.S. Patent 6,458,830, 2001. *Chem. Abstr.* 2000, 133, 252310x and 2000, 133, 266729c; (b) Nelson, T. D.; Rosen, J. D.; Smitroviych, J. H.; Payack, J.; Craig, B.; Matty, L.; Huffman, M. A.; McMamara, J. Org. Lett. 2005, 7, 55; (c) Dirat, O.; Elliott, J. M.; Jelly, R. A.; Jones, A. B.; Reader, M. Tetrahedron Lett. 2006, 47, 1295.
- Elliott, J. M.; Carlson, E. J.; Chicchi, G. G.; Dirat, O.; Dominguez, M.; Gerhard, U.; Jelly, R.; Jones, A. B.; Kurtz, M. M.; Tsao, K. L.; Wheeldon, A. *Bioorg. Med. Chem. Lett.* 2006, 16, 2929.